Stereoselective Synthesis of Homoneryl and Homogeranyl Triazole Bisphosphonates

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Supporting Information

ABSTRACT: Isoprenoid-substituted bisphosphonates are known to serve as inhibitors of the enzyme geranylgeranyl diphosphate synthase, and their activity can be highly sensitive to olefin stereochemistry. A mixture of homogeranyl and homoneryl triazole bisphosphonates has previously demon-



strated potent activity, and thus stereocontrolled syntheses of the individual isomers have been developed.

I nhibitors of the enzyme farnesyl diphosphate synthase (FDPS), including pamidronate (1) and zoledronate (2), are in clinical use for treatment of osteoporosis and diseases of the bone such as Paget's disease¹ and have become the standard of care for patients with multiple myeloma bone disease (Figure 1).² Inhibition of FDPS leads to depletion of the isoprenoid





farnesyl diphosphate (FDP) as well as the downstream product geranylgeranyl diphosphate (GGDP), and there is evidence that these agents exert their pharmacological effects by virtue of the depletion of GGDP. The C_{20} isoprenoid GGDP is formed from the C_{15} FDP and the C_5 isopentenyl diphosphate (IPP) in a reaction mediated by the enzyme geranylgeranyl diphosphate synthase (GGDPS).³ If depletion of GGDP is the proximate cause of the biological activity of the nitrogenous bisphosphonates, then inhibition of GGDPS may represent a more direct way to achieve the desired biological effect.

Our earliest studies of GGDPS inhibition identified several isoprenoid bisphosphonates that affect this enzyme selectively, perhaps most notably digeranyl bisphosphonate (3, DGBP, Figure 2) that has an IC_{50} of approximately 260 nM against the isolated enzyme.⁴ Several other dialkyl bisphosphonates have been reported to show comparable activity,^{5,6} and similar activity has been found more recently in the closely related ether 4.⁷ While further improvements on the V-shaped motif of these compounds may yet be possible,⁸ in our recent efforts^{9,10} to secure more potent inhibitors we have examined bisphosphonates assembled through click chemistry.^{11,12} In this vein, the most potent inhibitor we have identified to date is the bisphosphonate 5.¹³ This material is readily available via



Figure 2. Bisphosphonate GGDPS inhibitors and the initial retrosynthesis of the most potent example (5).

cycloaddition of the acetylene 6 and the azide 7, with the azide prepared from the alcohol 8 which in turn is readily available through a ring opening rearrangement of the cyclopropyl carbinol 9.¹⁴ Unfortunately, this rearrangement is known to give a mixture of olefin isomers in a ratio of $\sim 3:1$ (E to Z).¹⁴ Variations in the reaction conditions did improve the ratio somewhat in favor of the *E*-isomer, 15 but the homonerol isomer was of special interest because the nervl analogue already was known to be more active than the corresponding geranyl isomer.⁷ As the homoallylic bromides, the olefin isomers were not readily separable and so the mixture was carried forward to provide material for initial bioassays. Those efforts were rewarded when the olefin mixture 5 was found to have an IC₅₀ of 46 nM in enzyme assays,¹³ approximately 4-fold more potent than DGBP, and attractive activity in cellular assays as well. The increased potency of this mixture relative to earlier

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Scheme 1. Synthesis of the Homoneryl Isomer 20



Scheme 2. Synthesis of the Homogeranyl Isomer 29



compounds encouraged studies of the individual olefin isomers. In this report we describe the syntheses of both the E- and Z- olefin isomers of compound **5**.

Our initial efforts¹⁵ to prepare homonerol were based on use of the Wittig reagent prepared from the THP derivative of 3bromopropanol,¹⁶ which has been shown to be Z-selective in its reactions with aldehydes.¹⁷ However, while condensation of this reagent with commercial 6-methyl-5-hepten-2-one gave the THP derivative of compound 8 as a 1.4:1 mixture of Z- and Eolefin isomers, those isomers were not readily separated either as the THP acetals or as the free alcohols.¹⁵ There are several reports on preparation of homonerol and homogeraniol^{17–19} or the corresponding carboxylic acids,²⁰ but to secure pure samples of the individual isomers a new sequence derived from research reported by Wessjohann and co-workers²¹ appeared to be particularly attractive (Scheme 1). Their strategy relied upon classical acetoacetate chemistry and recently was employed in a clever synthesis of epothilone D^{22} For that effort, the Z stereochemistry of a trisubstituted olefin was derived from neryl bromide and cleanly preserved throughout their reaction sequence.

For our purposes, the β -keto acetate derivative **10** was prepared from neryl bromide and *tert*-butyl 2-acetoxy-acetoacetate according to the known procedure,²² and subsequent decarboxylation upon treatment with TsOH gave racemic acetate **11** (Scheme 1).²¹ Hydrolysis of this acetate proceeded smoothly to the acyloin **12**,²³ but then our efforts to bring about oxidative cleavage to the carboxylic acid went unrewarded. In contrast, reduction of compound **11** proceeded smoothly upon treatment with LiAlH₄ to give the diol **13** as a

mixture of stereoisomers, and oxidative cleavage by treatment with sodium periodate on silica gel gave the expected aldehyde 14 in nearly quantitative yield.

Once homoneral (14) was in hand, the remaining steps in the synthetic sequence proceeded as expected. Reduction of aldehyde 14 proceeded smoothly to give the homoallylic alcohol 15 as a single olefin isomer. After conversion of this alcohol to the corresponding mesylate (16) and a subsequent reaction with sodium azide to obtain the alkyl azide 17, the click reaction with acetylene 18 gave the desired triazole 19. Hydrolysis of the tetraester 19 under standard McKenna conditions²⁴ gave the final product 20, with no apparent formation of the isomeric olefin. The final product as well as the late stage intermediates were identified as single olefin isomers on the basis of their ¹³C NMR spectra. The C-4 methylene group of nerol and geraniol have significantly different resonances in their ¹³C NMR spectra (32.2 and 39.7 ppm, respectively),²⁵ and the corresponding C-5 methylene resonances for homonerol (32.2 ppm) and homogeraniol (40.0 ppm) are virtually identical. In compound 19, which was soluble in CDCl₃, this resonance was observed at 32.2 ppm; in compound 20, which was soluble in D_2O_1 , it was observed at 31.5 ppm. While the resonance of the α carbon in the bisphosphonate also is found in this range, it can be unambiguously identified by the large coupling constant with the two phosphorus atoms (\sim 120 Hz).

A parallel reaction sequence was employed to obtain the corresponding homogeranyl isomer (Scheme 2). In this series, the early reactions with the geranyl derivatives proceeded under parallel conditions and in nearly the same yields as those with

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the neryl isomer. For experimental convenience, the mesylate **26** again was converted directly to the azide **27** and then the azide was carried immediately into the click reaction with alkyne **18** to give the triazole **28** as the tetra ethyl ester. Hydrolysis then proceeded under parallel conditions to give the desired salt **29**. This sequence also gave a single olefin isomer throughout. For the ester **28**, the resonance for the key C-5 methylene group was observed at 39.8 ppm (CDCl₃) while the corresponding resonance in the salt **29** was found at 39.3 ppm (D₂O).

In conclusion, isomerically pure samples of both the Z- and E-olefin isomers **20** and **29** have been prepared through short synthetic sequences based on classic acetoacetate chemistry. These sequences can be conducted on a gram scale and have provided materials appropriate for further biological investigations. Based on the initial Western blot analyses, the homoneryl isomer does appear to be more potent as an inhibitor of GGDPS than the homogeranyl isomer, which is consistent with the relative activity of the neryl/geranyl pair,⁷ but more quantitative comparisons will require further studies.

EXPERIMENTAL SECTION

General Experimental Procedures. Diethyl ether was freshly distilled from sodium and benzophenone, whereas methylene chloride was distilled from calcium hydride prior to use. All other reagents and solvents were purchased from commercial sources and used without further purification. All reactions in nonaqueous solvents were conducted in flame-dried glassware under a positive pressure of argon and with a magnetic stir bar. NMR spectra were obtained at 300 MHz for ¹H, 75 or 125 MHz for ¹³C NMR, and 121 MHz for ³¹P NMR, in CDCl₃ with (CH₃)₄Si (¹H, 0.00 ppm) or CDCl₃ (¹H, 7.26 ppm; ¹³C NMR; 77.0 ppm) for non–aqueous samples or D₂O (¹H, 4.80 ppm) and 1,4-dioxane (¹³C, 67.19) for aqueous samples, as the internal standards. High resolution mass spectra were obtained by GC-TOF.

(5Z)-3-Acetoxy-6,10-dimethyl-5,9-undecadien-2-one (11). According to the published procedure,²¹ p-TsOH·H₂O (260 mg, 1.37 mmol) was added to a stirred solution of β -keto ester 10 (4.69 g, 13.3 mmol) in benzene (40 mL) at room temperature. The resulting solution was heated at 78 °C for 90 min and then was allowed to stir for 2 days at room temperature. The reaction mixture then was filtered through a bed of silica, the silica was rinsed with EtOAc, and the combined filtrate was concentrated *in vacuo* to afford the desired keto acetate 11 (3.34 g, 99%) as an orange oil which was used without further purification. The ¹H and ¹³C NMR data were consistent with the literature data.²¹

(5Z)-2,3-Dihydroxy-6,10-dimethyl-5,9-undecadiene (13). LiAlH₄ (247 mg, 6.51 mmol) was added to a stirred solution of keto acetate 11 (912 mg, 3.61 mmol) in Et₂O (18 mL) at 0 °C. After it was stirred for 2 h and allowed to warm slowly, the reaction then was cooled to 0 °C, quenched by slow addition of 1 N HCl followed by H₂O, and extracted with Et_2O (3×). The combined organic extracts were dried (Na_2SO_4) and filtered, and the filtrate was concentrated in vacuo to afford the desired diol 13 (685 mg, 89%) as a yellow oil. This mixture of diastereomers was used in the next step without further purification: ¹H NMR (300 MHz, CDCl₃) δ 5.23–5.05 (m, 2H), 3.89–3.31 (m, 2H), 2.28-2.16 (m, 2H), 2.12-2.04 (m, 4H), 2.02-1.98 (m, 1H), 1.94–1.88 (m, 1H), 1.74 (d, J = 1.2 Hz, 3H), 1.68 (s, 3H), 1.61 (d, J = 1.2 Hz, 3H), 1.22-1.16 (m, 3H); ¹³C NMR (75 MHz, CDCl₃) for the major isomer δ 137.8, 131.6, 124.1, 120.7, 76.1, 70.3, 32.1, 26.5, 25.7, 23.5, 19.4, 17.6, 16.6; HRMS (ES⁺) m/z calcd for C₁₃H₂₄O₂Na (M + Na)+ 235.1674, found 235.1679.

(Z)-4,8-Dimethylnona-3,7-dienal (14). In a manner similar to the published procedure,²⁶ sodium periodate (2.57 g, 12.03 mmol) was dissolved in hot water (5 mL, 70 °C), and silica gel (10.04 g) was added, followed by vigorous shaking to yield sodium periodate coated silica gel. The prepared periodate reagent (510 mg, 0.35 mmol) was

suspended in CH_2Cl_2 (2.7 mL) and allowed to stir for 10 min. To the suspension was added diol **13** (51 mg, 0.24 mmol), and the mixture was allowed to stir for 30 min, followed by filtration of the reaction mixture through diatomaceous earth, and removal of solvent using a rotary evaporator to yield the desired aldehyde **14** (39 mg, 100%) as a yellow oil that was used without further purification. The ¹H and ¹³C NMR data were consistent with the literature data.²⁷

(Z)-4,8-Dimethylnona-3,7-dien-1-ol (15). To a stirred suspension of LiAlH₄ (95%, 93 mg, 2.44 mmol) in Et₂O (27 mL) at 0 °C, aldehyde 14 (677 mg, 4.07 mmol) was added as a solution in Et₂O (1 mL) over 4 min. The reaction was allowed to warm to room temperature and stirred overnight. The reaction was cooled to 0 °C, quenched by slow addition of 1 N HCl followed by H₂O, and extracted with Et₂O. The combined organic extracts were dried (Na₂SO₄) and filtered, and the filtrate was concentrated *in vacuo*. Final purification by column chromatography (5% EtOAc in hexanes) afforded the desired alcohol 15 (654 mg, 95%) as a clear oil with a ¹H NMR spectrum in agreement with known data;^{28 13}C NMR (75 MHz, CDCl₃) δ 139.1, 132.1, 124.3, 121.0, 62.8, 32.2, 31.6, 26.8, 25.9, 23.7, 17.9.

(Z)-4,8-Dimethylnona-3,7-dien-1-yl Methanesulfonate (16). In a manner similar to a published procedure,²⁹ NEt₃ (0.25 mL, 1.78 mmol) was added to a stirred solution of homonerol (15, 178 mg, 1.06 mmol) in CH₂Cl₂ (10 mL) at 0 °C. After the reaction was allowed to stir at 0 °C for 20 min, methanesulfonyl chloride (0.13 mL, 1.68 mmol) was added dropwise to the reaction mixture. The reaction was allowed to stir for 3 h at 0 °C, followed by addition of H₂O to quench the reaction mixture. The resulting mixture was washed with 1 N HCl (2×), brine (1×), and NaHCO₃ (2×). The combined organic extracts were dried (Na₂SO₄) and filtered, and the filtrate was concentrated *in vacuo* to afford desired mesylate 16 (247 mg, 95%) as a yellow oil that was carried immediately to the next step.

Tetraethyl (3Z)-(2-(1-(4,8-Dimethylnon-3,7-dien-1-yl)-1H-1,2,3*triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate)* (19). In a manner similar to published procedures,^{11,30} NaN₃ (473 mg, 7.28 mmol) was added to a stirred solution of homoneryl mesylate (16, 944 mg, 3.83 mmol) in DMF (20 mL) under an argon atmosphere. The mixture was heated to 40 °C and allowed to stir overnight. The reaction then was diluted with Et_2O_1 , washed with water (5×) and brine, dried (Na₂SO₄), and filtered, and the filtrate was concentrated in vacuo, to yield homoneryl azide (17, 626 mg, 84%) as a yellow oil. The material was used immediately in the following reaction without further purification. To a stirred solution of tetraethyl but-3-yne-1,1diyldiphosphonate³¹ (18, 770 mg, 2.36 mmol) and homoneryl azide (17, 616 mg, 3.19 mmol) in t-BuOH/H₂O (4:1, 25 mL total), saturated CuSO₄ (0.01 mL) and sodium ascorbate (140 mg, 0.71 mmol) were added in sequence. The resulting reaction mixture was allowed to stir overnight at room temperature, and then the solvent was removed in vacuo. The resulting residue was dissolved in brine and extracted with EtOAc $(5\times)$. The combined organic extracts were washed with 5% NH₄OH, dried (Na₂SO₄), and filtered, and the filtrate was concentrated in vacuo. Final purification by column chromatography (10% EtOH in hexanes) afforded triazole 19 (982 mg, 80%) as a yellow oil: ¹H NMR (300 MHz, CDCl₃) δ 7.46 (s, 1H), 5.12–5.03 (m, 2H), 4.26 (t, J = 7.5 Hz, 2H), 4.19–4.06 (m, 8H), 3.39–3.24 (td, $J_{HP} = 16.1$ Hz, J = 6.4 Hz, 2H), 3.06–2.85 (m, 1H), 2.60–2.51 (m, 2H), 2.04-1.96 (m, 4H), 1.69 (s, 3H), 1.67 (s, 3H), 1.59 (s, 3H), 1.32–1.36 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 145.2 (t, J_{CP} = 8.8 Hz), 139.6, 132.2, 124.0, 122.4, 119.6, 63.0 (d, J_{CP} = 6.6 Hz, 2C), 62.7 (d, J_{CP} = 6.4 Hz, 2C), 50.5, 36.9 (t, J_{CP} = 133.0 Hz), 32.2, 29.2, 26.5, 25.9, 23.6, 22.3 (t, J_{CP} = 4.9 Hz), 17.9, 16.6 (d, J_{CP} = 3.7 Hz, 2C), 16.6 (d, J_{CP} = 3.4 Hz, 2C); ³¹P NMR (121 MHz, CDCl₃) 22.5 ppm; HRMS (ES⁺) m/z calcd for C₂₃H₄₄N₃O₆P₂ (M + H)⁺ 520.2705, found 520.2698.

Sodium (3Z)-(2-(1-(4,8-Dimethylnon-3,7-dien-1-yl)-1H-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (**20**). In a manner similar to published procedures, 9,24 to a stirred solution of bisphosphonate ester **19** (938 mg, 1.81 mmol) in CH₂Cl₂ (30 mL) at 0 °C, collidine (1.67 mL, 12.6 mmol) and TMSBr (97%, 1.97 mL, 15.2 mmol) were added dropwise in succession. The reaction was allowed to stir

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overnight while it warmed to room temperature, and the solvent then was removed in vacuo. The resulting residue was diluted with toluene (40 mL) and concentrated in vacuo to remove any excess TMSBr (3×). It then was treated with 2 N NaOH (6.05 mL, 12.1 mmol) and allowed to stir overnight at room temperature. Anhydrous acetone was added, and the mixture was placed in the freezer for 20 min. The resulting solid was collected by filtration, dissolved in water, and reprecipitated by addition of anhydrous acetone, and the mixture was placed in the freezer for 20 min. The resulting solid was collected by filtration, dissolved in water, and lyophilized to provide the desired salt 20 (501 mg, 56%) as a white powder: ¹H NMR (300 MHz, D₂O) δ 7.84 (s, 1H), 5.21–5.10 (m, 2H), 4.39 (t, J = 6.6 Hz, 2H), 3.21 (td, J_{HP} = 15.2 Hz, I = 6.6 Hz, 2H + 2.63 - 2.55 (m, 2H), 2.18 - 1.86 (m, 5H), 2.181.68 (m, 6H), 1.61 (s, 3H); ¹³C NMR (125 MHz, D₂O) δ 147.5, 140.8, 134.3, 124.8, 124.7, 120.6, 50.9, 40.1 (t, $J_{CP} = 116.7$ Hz), 31.5, 28.9, 26.3, 25.5, 23.1, 22.3 (t, $J_{CP} = 4.0$ Hz), 17.5; ³¹P NMR (121 MHz, D₂O) 18.7 ppm; HRMS (ES⁻) m/z calcd for C₁₅H₂₆N₃O₆P₂ (M -H)⁻ 406.1297, found 406.1289.

(5*E*)-2,3-*Dihydroxy*-6,10-*dimethyl*-5,9-*undecadiene* (23). According to the procedure for preparation of diol 13, keto ester 22 (1.26 g, 5.00 mmol) was treated with LiAlH₄ (595 mg, 14.9 mmol) in Et₂O (34 mL) at 0 °C. A parallel workup afforded the desired diol 23 (1.04 g, 98%) as a yellow oil that was used without further purification: ¹H NMR (300 MHz, CDCl₃) δ 5.20–5.12 (m, 1H), 5.10–5.01 (m, 1H), 3.90–3.76 (m, 1H), 3.69–3.54 (m, 1H), 2.10–1.96 (m, 6H), 1.68 (s, 3H), 1.64 (s, 3H), 1.60 (s, 3H), 1.25 (br s 3H); ¹³C NMR (75 MHz, CDCl₃) δ 139.7, 132.1, 124.3, 113.2, 75.9, 70.6, 40.0, 29.9, 26.7, 25.9, 19.5, 17.9, 17.4; HRMS (ES⁺) *m/z* calcd for C₁₃H₂₄O₂Na (M + Na)⁺ 235.1674, found 235.1658.

(E)-4,8-Dimethylnona-3,7-dienal (24). According to the procedure for the preparation of aldehyde 14, diol 23 (1.78 g, 8.38 mmol) was added to a stirred suspension of periodate coated silica gel (17.67 g, 12.1 mmol) in CH_2Cl_2 (56 mL). A parallel workup afforded the desired aldehyde 24 (1.39 g, 100%) as a yellow oil that was used without further purification. The ¹H NMR data were consistent with the literature data.²⁷

(E)-4,8-Dimethylnona-3,7-dien-1-ol (25). According to the procedure for the preparation of homoallylic alcohol 15, aldehyde 24 (1.39 g, 8.38 mmol) was added to a suspension of LiAlH₄ (180 mg, 5.03 mmol) in Et₂O (55 mL) at 0 °C. A parallel workup afforded homogeraniol (25, 1.35 g, 96%) as a yellow oil that was used without further purification. The ¹H NMR data were consistent with the literature data for material prepared by a different method.^{32,32}

(E)-4,8-Dimethylnona-3,7-dien-1-yl Methanesulfonate (26). According to the procedure for the preparation of mesylate 16, homogeraniol (25, 419 mg, 2.49 mmol) was treated with NEt₃ (0.59 mL, 4.20 mmol) followed by MsCl (0.31 mL, 4.01 mmol) in CH₂Cl₂ (25 mL) at 0 °C. A parallel workup afforded the desired mesylate 26 (542 mg, 88%) as a yellow oil that was used immediately without further purification.

Tetraethyl (3E)-(2-(1-(4,8-Dimethylnon-3,7-dien-1-yl)-1H-1,2,3triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (28). According to the procedure for the preparation of triazole 19, mesylate 26 (161 mg, 0.65 mmol) was treated with NaN₃ (80.1 mg, 1.23 mmol), and the resulting azide (27, 111 mg, 0.57 mmol) was isolated and immediately treated with acetylene bisphosphonate 18 (109 mg, 0.33 mmol) saturated CuSO₄ (0.01 mL), and sodium ascorbate (25 mg, 0.13 mmol) in sequence. A parallel workup and purification afforded the desired triazole 28 (112 mg, 65%) as a yellow oil: ¹H NMR (300 MHz, CDCl₃) δ 7.40 (s, 1H), 5.06–4.95 (m, 2H), 4.24–4.18 (t, J = 7.3 Hz, 2H), 4.12–4.04 (m, 8H), 3.32–3.19 (td, J_{HP} = 16.2 Hz, J = 6.5 Hz, 2H), 3.00-2.80 (tt, $J_{HP} = 23.5$ Hz, J = 6.3 Hz, 1H), 2.54-2.46 (dt, J = 7.9 Hz, 7.3 Hz, 2H), 2.00–1.88 (m, 4H), 1.61 (s, 3H), 1.53 (s, 3H), 1.48 (s, 3H), 1.26–1.19 (m, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 145.0, 139.7, 131.9, 124.1, 122.5, 118.8, 63.1 (d, J_{CP} = 6.5 Hz, 2C), 62.8 (d, J_{CP} = 6.5 Hz, 2C), 50.3, 39.8, 36.8, 29.4, 26.7, 25.9, 22.3 (t, J_{CP} = 4.9 Hz), 17.9, 16.5 (d, J_{CP} = 3.4 Hz, 2C), 16.4 (d, J_{CP} = 3.7 Hz, 2C), 16.3; ³¹P NMR (121 MHz, CDCl₃) 22.5 ppm; HRMS (ES⁺) m/z calcd for $C_{23}H_{44}N_3O_6P_2$ (M + H)⁺ 520.2705, found 520.2703.

Sodium (3*E*)-(2-(1-(4,8-Dimethylnon-3,7-dien-1-yl)-1*H*-1,2,3-triazol-4-yl)ethane-1,1-diyl)bis-(phosphonate) (**29**). According to the procedure for the preparation of sodium salt **20**, the bisphosphonate ester **28** (1.10 g, 2.12 mmol) was treated with TMSBr (97%, 2.30 mL, 17.7 mmol), collidine (1.95 mL, 14.6 mmol), and then 2 N NaOH (7.2 mL). A parallel workup and precipitation provided the desired sodium salt **29** (1.05 g, 70%) as a white powder: ¹H NMR (300 MHz, D₂O) δ 7.71 (s, 1H), 5.08–4.98 (m, 2H), 4.27–4.20 (t, *J* = 6.8 Hz, 2H), 3.10–2.95 (td, *J*_{HP} = 15.0 Hz, *J* = 6.8 Hz, 2H), 2.52–2.42 (m, 2H), 2.10–1.80 (m, 5H), 1.56 (s, 3H), 1.48 (s, 3H), 1.36 (s, 3H); ¹³C NMR (75 MHz, D₂O) δ 150.5 (t, *J*_{CP} = 7.3 Hz), 140.4, 134.0, 124.8, 124.4, 119.6, 50.5, 41.8 (t, *J*_{CP} = 118.1 Hz), 39.3, 28.8, 26.2, 25.4, 24.3, 17.5, 15.6; ³¹P NMR (121 MHz, D₂O) 18.7 ppm; HRMS (ES⁻) *m/z* calcd for C₁₅H₂₆N₃O₆P₂ (M – H)⁻ 406.1297, found 406.1304.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b01693.

The ¹H and ¹³C NMR spectra of compounds 13, 19, 20, 23, 28, and 29 and the ¹H NMR spectra of compounds 15 and 25 (PDF)

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Notes

The authors declare the following competing financial interest(s): D.F.W. is a named inventor of intellectual property related to digeranyl bisphosphonate that is owned by the University of Iowa Research Foundation. He is a founder of Terpenoid Therapeutics, Inc., which has licensed this property.

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REFERENCES

(1) Ebetino, F. H.; Hogan, A. M. L.; Sun, S.; Tsoumpra, M. K.; Duan, X.; Triffitt, J. T.; Kwaasi, A. A.; Dunford, J. E.; Barnett, B. L.; Oppermann, U.; Lundy, M. W.; Boyde, A.; Kashemirov, B. A.; McKenna, C. E.; Russell, R. G. G. *Bone* **2011**, *49*, 20.

(3) Wiemer, A. J.; Wiemer, D. F.; Hohl, R. J. Clin. Pharmacol. Ther. 2011, 90, 804.

(4) Wiemer, A. J.; Yu, J. S.; Lamb, K. M.; Hohl, R. J.; Wiemer, D. F. Bioorg. Med. Chem. 2008, 16, 390.

(5) Chen, C. K. M.; Hudock, M. P.; Zhang, Y. H.; Guo, R. T.; Cao, R.; No, J. H.; Liang, P. H.; Ko, T. P.; Chang, T. H.; Chang, S.; Song, Y. C.; Axelson, J.; Kumar, A.; Wang, A. H. J.; Oldfield, E. *J. Med. Chem.* **2008**, *51*, 5594.

(6) Barney, R. J.; Wasko, B. M.; Dudakovic, A.; Hohl, R. J.; Wiemer, D. F. Bioorg. Med. Chem. **2010**, *18*, 7212.

(7) Zhou, X.; Reilly, J. E.; Loerch, K. A.; Hohl, R. J.; Wiemer, D. F. Beilstein J. Org. Chem. 2014, 10, 1645.

(8) Foust, B. J.; Allen, C.; Holstein, S. A.; Wiemer, D. F. Bioorg. Med. Chem. 2016, 24, 3734.

⁽²⁾ Terpos, E.; Roodman, G. D.; Dimopoulos, M. A. Blood 2013, 121, 3325.

The Journal of Organic Chemistry

- (9) Zhou, X.; Ferree, S. D.; Wills, V. S.; Born, E. J.; Tong, H.; Wiemer, D. F.; Holstein, S. A. *Bioorg. Med. Chem.* **2014**, *22*, 2791.
- (10) Zhou, X.; Hartman, S. V.; Born, E. J.; Smits, J. P.; Holstein, S.
- A.; Wiemer, D. F. *Bioorg. Med. Chem. Lett.* **2013**, *23*, *764*. (11) Skarpos, H.; Osipov, S. N.; Vorob'eva, D. V.; Odinets, I. L.;
- Lork, E.; Roschenthaler, G. V. Org. Biomol. Chem. 2007, 5, 2361. (12) Feldman, A. K.; Colasson, B.; Sharpless, K. B.; Fokin, V. V. J.
- Am. Chem. Soc. 2005, 127, 13444.
 (13) Wills, V. S.; Allen, C.; Holstein, S. A.; Wiemer, D. F. ACS Med. Chem. Lett. 2015, 6, 1195.
- (14) McCormick, J. P.; Barton, D. L. J. Org. Chem. **1980**, 45, 2566.
- (15) Wills, V. S. Ph.D., University of Iowa, 2015.
- (16) Schow, S. R.; McMorris, T. C. J. Org. Chem. 1979, 44, 3760.
- (17) Mukai, C.; Ohta, M.; Yamashita, H.; Kitagaki, S. J. Org. Chem.
- **2004**, 69, 6867.
- (18) Yanagisawa, A.; Habaue, S.; Yasue, K.; Yamamoto, H. J. Am. Chem. Soc. 1994, 116, 6130.
- (19) Yanagisawa, A.; Yasue, K.; Yamamoto, H. Org. Synth. 1997, 74, 178.
- (20) Moragas, T.; Cornella, J.; Martin, R. J. Am. Chem. Soc. 2014, 136, 17702.
- (21) Scheid, G.; Kuit, W.; Ruijter, E.; Orru, R. V. A.; Henke, E.; Bornscheuer, U.; Wessjohann, L. A. *Eur. J. Org. Chem.* 2004, 2004, 1063.
- (22) Wessjohann, L. A.; Scheid, G. O.; Eichelberger, U.; Umbreen, S. J. Org. Chem. 2013, 78, 10588.
- (23) Scheid, G.; Ruijter, E.; Konarzycka-Bessler, M.; Bornscheuer, U. T.; Wessjohann, L. A. *Tetrahedron: Asymmetry* **2004**, *15*, 2861.
- (24) McKenna, C. E.; Higa, M. T.; Cheung, N. H.; McKenna, M. C. Tetrahedron Lett. **1977**, *18*, 155.
- (25) Bohlmann, F.; Zeisberg, R.; Klein, E. Org. Magn. Reson. 1975, 7, 426.
- (26) Zhong, Y.-L.; Shing, T. K. M. J. Org. Chem. 1997, 62, 2622.
- (27) Baillargeon, V. P.; Stille, J. K. J. Am. Chem. Soc. **1986**, 108, 452.
- (28) Schulteelt, K. H.; Snowden, R. L.; Tarchini, C.; Baer, B.; Vial, C.; Schulte-Elte, K. H. U.S. Patent #5077417A, December 31, 1991.
- (29) Gash, R. C.; Maccorquodale, F.; Walton, J. C. Tetrahedron 1989, 45, 5531.
- (30) Rebek, J.; Shaber, S. H.; Shue, Y. K.; Gehret, J. C.; Zimmerman, S. J. Org. Chem. **1984**, *49*, 5164.
- (31) Weinhart, M.; Groger, D.; Enders, S.; Dernedde, J.; Haag, R. Biomacromolecules **2011**, *12*, 2502.
- (32) Kocienski, P.; Wadman, S.; Cooper, K. J. Org. Chem. 1989, 54, 1215.